Modeling of gene regulation by DNA supercoiling in bacteria

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- Extensive regulation by DNA topology *i.e.* supercoiling (SC)?

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- Deformation of DNA double-helix resulting from torsional stress
- Constant regulation by **topoisomerases** which are **essential** and **highly conserved**



• A rapid physical sensor of changing environments in bacteria



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• Contribution to global gene expression reprogramming besides stress-specific response pathways?

• A global, complex (and ubiquitous?) regulator in bacteria

Gene response to DNA relaxation



• A global, complex (and ubiquitous?) regulator in bacteria

Phylum	Species	SC change	Method	Genes significantly affected (% genome)
Proteobacteria	Escherichia coli	Rel (+)	Norfloxacin	613 (15%)
		Rel (+)	Novobiocin / pefloxacin	1957 (48%)
		Rel (+)	Genetic engineering	740 (18 %)
		Rel (+)	Genetic engineering / norfloxacin / novobiocin	306 (7%)
	Salmonella typhimurium	Rel (+)	Genetic engineering	499 (10%)
	Dickeya dadantii	Rel (+)	Novobiocin	1461 (32%)
		Rel (+)		1212 (27%)
	Haemophilus influenzae	Rel (+)	Novobiocin / ciprofloxacin	640 (37 %)
Firmicutes	Streptococcus pneumoniae	Rel (+)	Novobiocin	290 (14.2%)
		Hyp (-)	Seconeolitsin	545 (27%)
	Staphylococcus aureus	Rel (+)	Novobiocin	280 (11 %)
	Bacillus subtilis	Rel (+)		1075 (24 %)
Actinobacteria	Streptomyces coelicolor	Rel (+)	Novobiocin	121 (1.5 %)
	Mycobacterium tuberculosis	Rel (+)		Not provided
Tenericutes	Mycoplasma pneumoniae	Rel (+)	Novobiocin	469 (43 %)
Cyanobacteria	Synechocystis	Rel (+)	Novobiocin	Several genes
	Synechococcus elongatus	Rel (+)		Not provided

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\Rightarrow No quantitative nor qualitative modeling

Modeling of gene regulation by DNA supercoiling



Complexity of transcriptomic response to σ variation

Modeling of gene regulation by DNA supercoiling



Complexity of transcriptomic response to σ variation

Hypothesis : promoter-specific response

Regulation without regulators

• Basal regulation of RNA Polymerase-DNA interaction during transcription initiation





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$$k(\sigma, seq) = k_0 \exp\left(\frac{\Delta G_{op}(\sigma, seq) + \Delta G_{RNAP}}{k_B T}\right)$$
$$\Delta G_{RNAP} = 3.5k_B T$$



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• Gene response to novobiocin-induced relaxation consistent with predictions?



• pheP hybrid strain in LB rich medium



• exp. : time point used to compare the expression variation among promoters

• Relaxation response of promoters with mutated discriminators match the predictions



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 \Rightarrow Selective repression depending on discriminator sequences \Rightarrow Global regulation mode ?

Analysis of whole-genome expression data

- Transcriptome under DNA relaxation
- Map of all promoters with sigma factors, discriminator and genes

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Predictions and confrontation to experimental data

$$k(\sigma, seq) = k_0 \exp\left(\frac{\Delta G_{op}(\sigma, seq) + \Delta G_{RNAP}}{k_B T}\right)$$

- Expression rates before/after DNA relaxation
- Normalization step to mimic transcriptomic protocol

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An ubiquitous regulation mode?



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- Contribution to the entire genomic response to relaxation?
- Proportion of accurately predicted promoters (sensitivity) and comparison to a random model \Rightarrow up to 15% predictability gain

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<u>Limitations</u> : promoters annotated with poor resolution, \neq drug and protocol used, local SC levels \neq global SC level of the chromosome, gene regulation at \neq transcription steps









- Up to 10% predictability gain
- The selective activation/repression of promoters by stress conditions is controlled by changes in DNA SC depending on discriminator sequences

14 / 16

Conclusion

• DNA Supercoiling, by modulating RNA Polymerase-DNA interaction, constitutes a global, complex and ubiquitous mode of gene regulation, based on fundamental mechanical properties of DNA

In eukaryotes?

- Open-complex formation : (1) ATP-dependent melting by TFIIH (optional?) (2) local negative DNA supercoiling provided by transcription or nucleosomes
- Similarities among eukaryotic and bacterial promoters in spite of evolutionary distances



CRP team, MAP lab, INSA Lyon

Merci pour votre attention

